

# Bio-control by using Antagonistic (Filamentous Fungi and VAM) and Bacteria against *Macrophomina phaseolina*

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## ABSTRACT

The excessive use of pesticides and fertilizers in the current agricultural system, which is done to increase production, completely eradicate plant pathogens, and reduce undesirable weeds, has a detrimental effect on soil quality, water body environment, animal and human health due to the toxicity, recalcitrance, and carcinogenic potential of many of these compounds. It has long been thought that biological plant disease control could replace current methods of prevention. Filamentous fungi (especially *Aspergillus* spp., *Trichoderma reesei*, and *Neurospora crassa*), bacteria (*Pseudomonas*, *Bacillus*, *Burkholderia*, *Lysobacter*, *Serratia*, and *Pantoea*), and Vesicular Arbuscular Mycorrhiza (VAM) (*G. mosseae*, *Glomus claroideum*, *Glomus aggregatum*) are employed instead of chemicals in biocontrol. *M. phaseolina* is a fungus that lives in the root soil and produces dry root rot/stem canker, stalk rot, and charcoal rot. The fungus *M. phaseolina* causes charcoal rot, stalk rot, and dry root rot/stem canker in plant roots. Melon, strawberries, and tomatoes are just a few of the horticultural crops that *M. phaseolina* has been discovered on throughout Europe, the US, Australia, Chile, and Israel. To manage *M. phaseolina*, various biocontrol agents (filamentous fungi, VAM, and bacteria) are used successfully and effectively.

**Keywords:** Alternative, bacteria, Antagonist fungi, Biocontrol, VAM, *M. phaseolina*.

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## INTRODUCTION

The demand for agricultural products and food grains rises along with the population. The deliberate application of chemical fertilizers and pesticides has greatly risen in the modern global agricultural system. More than two billion metric tons of pesticides are used annually throughout the world. The use of fungicides eliminates plant infections caused by fungi, bacteria, viruses, and nematodes, bactericides, herbicides, insecticides, and other pesticides (Dean *et al.*, 2012; Singh, 2014; O'Brien, 2017). Due to the toxicity, recalcitrance, and carcinogenic potential of many of these substances, excessive fertiliser use has a severe influence on soil quality, hydro-geological environment, both animal and human health. Pest biocontrol is now accepted as a practical alternative to pesticides. Around the world, fungi antagonists are used as biocontrol agents (BCAs) to protect plants from diseases and infections. As biocontrol agents, various antagonistic fungi (including filamentous fungus and VAM) and bacteria are used to prevent and treat diseases and enhance plant development and productivity.

## FILAMENTOUS FUNGI

Filamentous fungi (especially *Aspergillus* spp., *Trichoderma reesei*, and *Neurospora crassa*). It is well known that filamentous fungi secrete a large number of different hydrolytic enzymes, making them the main source of enzymes for the hydrolysis of lignocellulose (Stirling and Stirling, 1997). In the soil, filamentous fungi perform two key tasks: they break down organic materials and promote soil aggregation. Numerous fungi that are connected with plants are well known for promoting plant growth, and soil-borne microbes can enter roots and establish themselves as endophytes in plants (Sylvia *et al.*, 2005).

The antagonists can be specialized in a variety of ways to control biological ailments, both directly and indirectly (Fig. 1). These mechanisms include antibiosis (in which the antagonist

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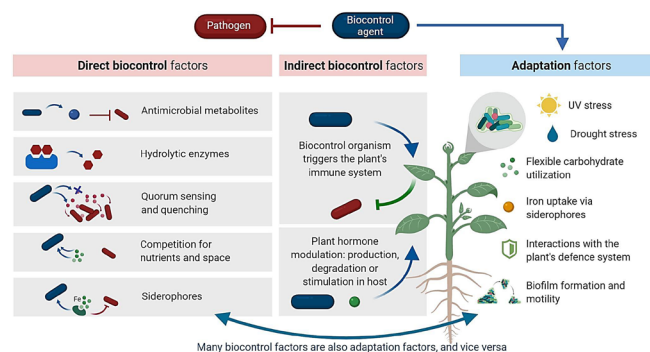
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produces an inhibitory metabolite or antibiotic), mycoparasitism (in which the antagonist obtains all or the majority of its nutrients from the fungal host), induced resistance (induction of plant defence responses against plant pathogens), and growth promotion (BCAs promote plant growth while the effects of the disease are being reduced and also through microbial hormones such as indole acetic acid and gibberellic acid). Further activities



**Fig. 1:** Interaction of Pathogens and biocontrol agents.

extracellular hydrolytic enzymes secreted by the antagonist, microbes competing for nutrients and space, and pathogenicity detoxification are a few biological disease control strategies (Zhang *et al.*, 2014; Deketelaere *et al.*, 2017). With the exception of oomycetes, which contain cellulose in their cell walls but no chitin, many plant pathogenic fungus have chitin and - (1, 3)-glucan as their two main structural constituents.

### Mechanism of Antagonist Fungi

It has been associated with the biological control of a number of soil-borne fungal infestations that microbes producing chitinases or glucanases exhibit antagonistic behavior toward fungus *in-vitro*. One of the few beneficial fungi that have been successfully used effectively as a biocontrol agent is the *Trichoderma* species. India, France, the UK, Switzerland, Sweden, Belgium, Chile, New Zealand, and the US have all recognized the microbe as a bio-fungicide; rules are still pending in a large number of additional nations.

*Trichoderma* often grows chemotropically in consequence to hyphal activation or gradients of the host's chemical production. The mycoparasite coils around the host or develops hook-like structures to adhere to it whenever it gets there. Sometimes during these interactions, the mycoparasite can enter the host mycelium by partially destroying it. The idea that *Trichoderma* spp. produce and secrete mycolytic enzymes that lead to a partial breakdown of the host's cell wall was based on microscopic findings. The six complementing modes of action of these enzymes, all of which may be necessary for maximal efficacy against chitin-containing plant pathogenic fungi, are what give rise to the appearance and diversity of the *T. harzianum* chitinolytic system.

### Antibiotic-mediated Suppression

Antibiotics are microbial toxins that, in small doses, can harm or kill other microbes. Bacteria can create both volatile and nonvolatile antibiotics, such as diacetyl phloroglucinol (DAPG) and mupirocin, as well as phenazine derivatives including pyocyanin, phenazine-1-carboxylic acid (PCA), PCN, and hydroxyphenazines (Ahmad *et al.*, 2008).

### Competition

When a biocontrol agent reduces a certain substance's availability and consequently restricts the pathogen's ability to develop, this is known as biocontrol *via* nutrient competition. In particular, the biocontrol agents are more effective at utilizing the chemical than the pathogens are at absorbing

it (Handelsman and Parke, 1989). Some bacteria, particularly *Fluorescent pseudomonads*, make siderophores with extremely high iron affinities that can snare this scarce resource from other microflora, inhibiting their growth.

### Parasitism

The direct use of one organism as food by another occurs during this phase (Handelsman and Parke, 1989). Mycoparasites are fungi that feed off of other fungi and are commonly seen (Baker and Cook, 1974.). On a variety of fungi, several mycoparasites exist, and some of them have been suggested to be crucial in the prevention and treatment of disease (Adams, 1990). Saccardo, for instance, characterized *Darluca filum* (now *Sphaerellopsis filum*) as a parasite of several rust fungi, particularly Puccinia and Uromyces (Sundheim and Tronsmo, 1988).

In general, *Trichoderma lignorum* (*T. viride*), which parasitizes *Rhizoctonia solani* hyphae, and suggestions of Mycoparasites can either coil around the host hyphae or grow beside it and release enzymes that attack the target fungus by dissolving its cell walls (Chet, 1987). These enzymes, such as chitinases and b-1,3-glucanase, may be involved in the breakdown of host cell walls and may be parts of complicated combinations of synergistic proteins that cooperate to combat pathogenic fungus (Di Pietro *et al.*, 1992). Penetration is the ultimate phase. To puncture the target fungus' cell wall, the biocontrol agent creates structures resembling appressoria (Chet, 1987). A specific BCA that kills the pathogen or its propagules directly attacks it in hyperparasitism.

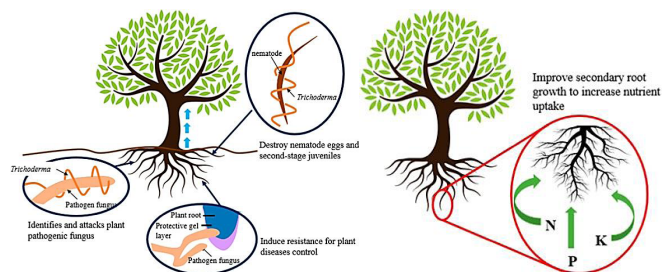
### Induction of Systemic Resistance

Systemic acquired resistance (SAR) is the term used to describe the induced resistance of plants to different diseases. Inoculating plants with a necrogenic pathogen, a nonpathogen, or specific natural or manufactured chemical substances can cause SAR (Lam and Gaffney, 1993). The synthesis of various proteins, such as chitinases, glucanases, peroxidases, and other pathogenesis-related (PR) proteins, as well as the physical thickening of cell walls through lignification, callose deposition, accumulation of antimicrobial low-molecular-weight substances (such as phytoalexins), are examples of these defence responses (Hammerschmidt *et al.*, 1984).

According to the host-parasite connection under consideration, different levels of hydrolytic enzymes are produced. This phenomenon is associated with the particular disease-controlling properties of each *Trichoderma* isolate. Mycoparasitism is considered to be one of the primary mechanisms underlying *Trichoderma*'s antagonistic behavior as



Fig. 2: Mechanism of Biocontrol agents.



(Nur and Badaluddin, 2020)

Fig. 3: Role of biocontrol agents in disease management and soil health improvement.

a biocontrol agent, along with chemotropic growth, the release of extracellular enzymes, and host lyses (Fig. 2). *Trichoderma*'s ability to biocontrol is most likely the result of several different processes (Howell, 2003). *In vitro* antibiotics that are effective against infections are produced by the well-known biological control agent *Trichoderma* (Howell, 2003). (Fig. 3).

### Fungal Biocontrol Agents

The use of *Trichoderma* is completely safe for both humans and animals. The five main species utilized in the biocontrol of plant diseases are *Trichoderma harzianum*, *T. koningii*, *T. longibrachiatum*, *T. pseudokoningii*, and *T. viride*. Mycoparasitism is a difficult process with numerous steps. *Trichoderma*'s connection with its host is distinctive. Numerous *Trichoderma* species have undergone significant research as biocontrol agents (Papavizas, 1985).

### Bacterial Biocontrol Agents

Bacterial competition for nutrients and space, the avoidance of pathogen colonization of host tissues, the production of crop disease resistance in plants, and the accumulation of toxic metabolites (enzymes, antibiotics, and volatile organic compounds) are all causes of bacterial antagonism. By working through one or more routes, it has been discovered that the bacterial species belonging to the genera *Pseudomonas*, *Bacillus*, *Burkholderia*, *Lysobacter*, *Serratia*, and *Pantoea* are effective against a range of plant infections. The rhizobacterial species

are effective biocontrol agents and plant growth stimulators, giving the treated plants a two-fold advantage (Table 1).

### VAM (Vesicular - Arbuscular Mycorrhiza)

Symbiotic connections develop between higher plants and mycorrhizal fungi. Endotrophic or vesicular - arbuscular mycorrhiza (VAM) and ectotrophic mycorrhiza are the two forms of mycorrhiza that have been identified. The arbuscle is a dichotomously branching structure that is thought to be the site of plant-fungus nutrient exchange. The fungal partner in VAM is restricted to the cells of the plant cortex, where it grows both within and outside the cells. As a fungal partner, soil doesn't seem to have a distinct existence of its own. Vesicular-arbuscular mycorrhizas can directly improve the uptake of vital nutrients like as phosphorus, copper, and iron. Therefore, some plants are protected from these elements' harmful effects *via* mycorrhizal connections. Additionally, ectotrophic mycorrhizas consume more phosphorus, and by mineralizing organic nitrogen, they increase the availability of nitrogen to plants. They may also help plants absorb more water from the soil and defend their plant hosts from disease and heavy metal damage.

In symbiotic relationships with mycorrhizal fungi, around 90% of all plants produce hyphae networks. Mycorrhizae mostly draw phosphorus and other minerals, such zinc and copper, from the soil. The plant root provides the fungus with nutrients like glucose. This advantageous contact is known as a mycorrhizal network (Smith, 1997). Mycorrhizal fungi are responsible for the improved growth of host plant species due to increased nutrient intake, production of compounds that promote growth, tolerance to salt and drought, and synergistic interactions with other helpful bacteria (Maeder *et al.*, 2002). Arbuscular mycorrhiza fungi can alter the soil's characteristics, promoting plant growth in both benign and adverse conditions (Mo *et al.*, 2016). Arbuscular mycorrhiza fungi increase plant development and alter the morphological, nutritional, and physiological traits of plants to increase their ability to withstand abiotic stresses (Shekoofeh *et al.*, 2012).

**Table 1:** Some bacterial biocontrol agents used against diseases

Biocontrol agents	Disease
<i>Agrobacterium radiobacter</i>	crown gall
<i>Pseudomonas fluorescens</i>	bacterial blotch, seedling diseases
<i>Trichoderma viride</i>	timber pathogens
<i>Trichoderma</i> spp.	root diseases
<i>Trichoderma harzianum</i>	root diseases, wood decay
<i>Gliocadium virens</i>	seedling diseases

**Table 2:** Management of *Macrophomina phaseolina* pathogen by antagonistic fungi (filamentous fungi and VAM) and bacteria

Biocontrol agent	Disease	References
<i>T. harzianum</i> + <i>G. mosseae</i>	Reduced root-rot in geranium plants caused by <i>Macrophomina phaseolina</i>	Haggag and Abd-El latif (2001)
<i>Pseudomonas aeruginosa</i> , <i>Bacillus subtilis</i> , and <i>Burkholderia</i> species	Seed dressing and soil drenching with bacterial formulations of reduced dry root rot incidence of mungbean and urdbean.	Satya <i>et al.</i> , 2011; Latha <i>et al.</i> (2017)
<i>Trichoderma</i> isolates	Mungbean dry root rot was reduced when cultivated in organic substrates such groundnut shell, coir pith, and press mud.	Raghuchander <i>et al.</i> (1993)
<i>Trichoderma viride</i> or <i>Trichoderma harzianum</i> with <i>Pseudomonas fluorescens</i> or <i>B. subtilis</i>	lower incidence of mungbean and urdbean dry root rot in greenhouse and outdoor settings.	Kumari <i>et al.</i> (2015); Shahid & Khan, 2016)
<i>T. harzianum</i> or <i>T. viride</i>	reduced dry root rot of mungbean and urdbean	Athira, (2017); Jamwal <i>et al.</i> (2016)
<i>Trichoderma</i> species, the arbuscular mycorrhizal species <i>Glomus claroideum</i>	reduced dry root rot of mungbean and	Chandra <i>et al.</i> (2007).
<i>Glomus aggregatum</i>	reduced dry root rot of urd bean in field trials	Chandra <i>et al.</i> (2007).
<i>T.harzianum</i> , <i>T. Polysporum</i> and <i>T. viride</i>	Soil application of talc-based formulation effectively controlled the eggplant's root rot ( <i>M. phaseolina</i> ) under field conditions.	Ramezani, (2008)



## Macrophomina Phaseolina

*Macrophomina Phaseolina* results in root decay (Tassi) a number of crop plants are affected by the disease, which has a considerable economic impact (Jana *et al.*, 2005). The fungus *M. phaseolina* causes charcoal rot, stalk rot, and dry root rot/stem canker in plant roots. Different physiological and ecological factors, such as heat, high temperatures, and low moisture content, may make it more common. In Australia (Gomez *et al.*, 2020), Chile (Sánchez *et al.*, 2013), Israel (Sánchez *et al.*, 2013), Europe and the United States (Baggio *et al.*, 2019), *M. phaseolina* has also been discovered on melon, strawberry, and tomato. (Chamorro and others, 2015a). Under drought conditions, it also threatens sunflower output in South Asia (Khan *et al.*, 2017) and Italy (Manici *et al.*, 1995).

The most prominent diseases that have an impact on sunflower plant growth are root rot and charcoal rot, both of which are caused by *Macrophomina phaseolina*. The biological control of soil-borne plant diseases and the stimulation of plant growth have both made extensive use of microorganism enrichment, conservation, and management. Although *Bacillus* and *Streptomyces* species have also been utilized, *Fluorescent pseudomonas* are the most frequently used bacteria for biological control and plant growth promotion. With soil-borne plant infections as well as pathogens on phylloplane, competition has been used as a biological control method. Increased growth response was observed in lettuce bean, cucumber, and pepper following the treatment of *Trichoderma* spp. in pot culture or field conditions. *Trichoderma* has the capacity to modify plant hormones and vitamins, which may have an impact on plant growth (Harman *et al.*, 2004). It has been proposed that control of plant infections and induced resistance as indirect mechanisms for *Trichoderma*-mediated plant growth promotion.

Antagonistic fungi (filamentous fungus and VAM) and bacteria are used to manage the *Macrophomina phaseolina* pathogen (Table 2).

## CONCLUSION

Overusing chemicals as pesticides, weed killers or to treat various plant diseases or increase productivity creates risks to the health of people, animals, plants, and the environment because of the residue problem. One ecofriendly alternative to fungicides is biocontrol agents (BCAs). In addition to bacteria, VAM is a biocontrol agent for filamentous fungi. Biocontrol-active microorganisms can work in a variety of ways to prevent fungus-related plant diseases, including hyperparasitism, predation, antibiosis, cross-protection, competition for location and resources, and generating resistance. Most biocontrol treatments are used to treat soil- and seed-borne fungal infections, like those that cause seed rot, damping-off, and root rot. Utilizing biological control is a viable technique for controlling plant diseases, boosting productivity, safeguarding the environment and biological resources, moving toward a sustainable agricultural system, and efficiently resolving issues with chemical hazards.

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## REFERENCES

- Adams, P. B. 1990. The potential of mycoparasites for biological control of plant diseases. *Annu. Rev. Phytopathol* 28:59-72
- Ahmad, F., Ahmad, I., Khan, M.S. 2008. Screening of free-living rhizospheric bacteria for their multiple plant growth promoting activities. *Microbial Res.* 163:173–181
- Alqarawi, A.A., Abdallah, E.F., Hashem, A. 2014. Alleviation of salt-induced adverse impact via mycorrhizal fungi in *Ephedra aphylla* Forsk. *Journal of Plant Interactions* 9(1):802-810.
- Athira, K. 2017. Efficacy of fungicide and bio-control agent against root rot of black gram (*Vigna mungo* L.) caused by *Macrophomina phaseolina* (Tassi) Goid. *International Journal of Current Microbiology and Applied Science* 6: 2601– 2607.
- Baggio, J.S., Cordova, L.G. & Peres, N.A. 2019. Sources of inoculum and survival of *Macrophomina phaseolina* in Florida strawberry fields. *Plant Disease* 103: 2417– 2424.
- Baker, K. F., and Cook, R. J. 1974. Biological Control of Plant Pathogens. *Am. Phytopathol. Soc.*, St. Paul, MN. 433 pp.
- Chamorro, M., Domínguez, P., Medina, J.J., Miranda, L., Soria, C., Romero, F. 2015a. Assessment of chemical and biosolarization treatments for the control of *Macrophomina phaseolina* in strawberries. *Scientia Horticulturae* 192: 361– 368.
- Chandra, S., Khare, V. and Kehri, H.K. 2007. Evaluation of arbuscular mycorrhizal fungi against *Macrophomina phaseolina* causing dry root rot of urdbean and mungbean. *Indian Phytopathology* 60: 42– 47.
- Chet, I. 1987. *Trichoderma* application, mode of action, and potential as biocontrol agent of soil-borne pathogenic fungi. In: Innovative Approaches to Plant Disease Control. I. Chet, ed., John Wiley, New York. 137-160.
- Dean, R., Van Kan, J. A., Pretorius, Z.A., Hammond-Kosack, K.E., Di Pietro, A., Spanu, P.D. 2012. The Top 10 fungal pathogens in molecular plant pathology. *Mol. Plant Pathol.* 13 (4): 414–430.
- Deketelaere, S., Tyvaert, L., Franca, S. C., Hofte, M. 2017. Desirable traits of a good biocontrol agent against *Verticillium* wilt. *Front. Microbiol.* 8: 1186. 10.3389/fmicb.2017.01186
- Di Pietro, A. 1993. Chitinolytic enzymes produced by *Trichoderma harzianum* : antifungal activity of purified endochitinase and chitobiosidase. *Phytopathol.* 83: 302-307.
- Gianinazzi, S., Bosatka, M. 2004. Inoculum of arbuscular mycorrhizal fungi for production systems: Science meets business. *Canadian Journal of Botany* 82: 64-71
- Gomez, A.O., De Faveri, J., Neal, J.M., Aitken, E.A.B. and Herrington, M.E. 2020. Response of strawberry cultivars inoculated with *Macrophomina phaseolina* in Australia. *International Journal of Fruit Science* 20: 164– 177.
- Haggag, W.M. and Abdel-latif, F.M. 2001. Interaction between vasculararbuscular mycorrhizae and antagonistic biocontrol microorganisms on controlling root-rot disease incidence of geranium plants. *Online Journal of Biological Sciences* 1(12):1147-1153.
- Hammerschmidt, R., Lampert, D. T. A., and Muldoon, E. P. 1984. Cell wall hydroxyproline enhancement and lignin deposition as an early event in the resistance of cucumber to *Cladosporium cucumerinum*. *Physiol. Plant Pathol.* 24: 43 - 47 .
- Handelsman, Jo. and Parke, J.L. 1989. Mechanisms in biocontrol of soilborne plant pathogens. In: Plant-Microbe Interactions, Molecular and Genetic Perspectives, T. Kosuge, and E. W. Nester, eds., McGraw-Hill, New York. 3: 27-61.
- Harman, G. E., Howell, C. R., Viterbo, A., Chet, I. and Lorito, M. 2004. *Trichoderma* species – Opportunistic, Avirulent Plant Symbionts. *Nature Reviews Microbiology* 2: 43–56.
- Howell, C. R. 2003. Mechanisms employed by *Trichoderma* species in the biological control of plant diseases: The history and evolution of current concepts. *Plant Disease* 8: 4–10.
- Jamwal, S., Jamwal, A., Reena, K.A., Isher, A. and Dutta, U. 2016. Management of root rot of urdbean (*Phaseolus mungo*) with *Trichoderma* spp. in rainfed areas of Jammu and Kathua districts. *The Bioscan* 11: 2947– 2951.

- Jana, T. K., T. R. Sharma and N. K. Singh. 2005. SSR-based detection of genetic variability in the charcoal kinase of *Trichoderma virens*, is involved in biocontrol properties and repression of conidiation, pathogenicity, pycnidium production and chlorate utilization. *Can. J. Bot.* 73: 1596–1603
- Khan, A.N., Shair, F., Malik, K., Hayat, Z., Khan, M.A., Hafeez, F.Y. 2017. Molecular identification and genetic characterization of *Macrophomina phaseolina* strains causing pathogenicity on sunflower and chickpea. *Frontiers in Microbiology* 8: 1309.
- Kumari, R., Shekhawati, K.S., Gupta, R. and Khokhar, M.K. 2015. Integrated management against root rot of mungbean (*Vignaradiata* L. Wilczek) incited by *Macrophomina phaseolina*. *Journal of Plant Pathology and Microbiology* 3: 1–5.
- Lam, S. T., and Gaffney, T. D. 1993. Biological activities of bacteria used in plant pathogen control. In: *Biotechnology in Plant Disease Control*. Chet, I. ed., John Wiley, New York. 291-320.
- Latha, P., Karthikeyan, M. and Rajeswari, E. 2017. Development of bioformulations for the management of blackgram dry root rot caused by *Rhizoctonia bataticola* (Taub Butler). *Advances in Research* 9: 1–12.
- Maeder, P., Fliessbach, A., Dubois, D., Gunst, L., Fried, P., Niggli, U. 2002. Soil fertility and biodiversity in organic farming. *Science* 296:1694-1697
- Manici, L.M., Caputo, F. and Cerato, C. 1995. Temperature responses of isolates of *Macrophomina phaseolina* from different climatic regions of sunflower production in Italy. *Plant Disease* 79: 834–838.
- Mo, Y., Wang, Y., Yang, R., Zheng, J., Liu, C., Li, H. 2016. Regulation of plant growth, photosynthesis, antioxidation and osmosis by an arbuscular mycorrhizal fungus in watermelon seedlings under well-watered and drought conditions. *Frontiers in Plant Science* 7: 644
- Nur, A.Z. and Badaluddin, A. 2020. Biological functions of *Trichoderma* spp. for agriculture applications. *Annals of Agricultural Sciences*. 65(2): 168-178.
- O'Brien, P. A. 2017. Biological control of plant diseases. *Australas. Plant Pathol.* 46 (4): 293–304.
- Papavizas, G.C. 1985. *Trichoderma* and *Gliocladium*: Biology, ecology and potential for biocontrol. *Annual Review of Phytopathology* 23: 23-54.
- Raghuchander, T., Samiyappan, R. and Arjunan, G. 1993. Biocontrol of *Macrophomina* root rot of mungbean. *Indian Phytopathology* 46: 379–382.
- Ramezani, H. 2008. Biological control of root-rot of eggplant caused by *Macrophomina phaseolina*. *American-Eurasia. J. Agric. & Environ. Sci.* 4(2): 218-220.
- Sánchez, S.M., Gambardella, J.L., Henríquez, J.L. and Díaz, I. 2013. First report of crown rot of strawberry caused by *Macrophomina phaseolina* in Chile. *Plant Disease* 97: 996.
- Satya, V.K., Vijayasamundeeswari, A., Paranidharan, V. and Velazhahan, R. 2011. *Burkholderia* sp. strain TNAU-1 for biological control of root rot in mungbean (*Vignara diata* L.) caused by *Macrophomina phaseolina*. *Journal of Plant Protection Research* 51: 273–278.
- Shahid, S. and Khan, M.R. 2016. Biological control of root rot of mung bean plants incited by *M. phaseolina* through microbial antagonists. *Plant Pathology Journal*, 15: 27–39.
- Shekoofeh, E., Sepideh, H., Roya, R. 2012. Role of mycorrhizal fungi and salicylic acid in salinity tolerance of *Ocimum basilicum* resistance to salinity. *Journal of Biotechnology* 11(9): 2223-2235
- Singh, H. B. 2014. Management of plant pathogens with microorganisms. *Proc. Indian Natl. Sci. Acad.* 80 (2): 443–454.
- Smith, S.E., Read, D.J. 1997. *Mycorrhizal Symbiosis*. 2nd ed. London, UK: Academic Press. 605p
- Sreenivasa, M.N., Bagyaraj, D.J. 1989. Use of pesticides for mass production of vesicular arbuscular mycorrhizal inoculum. *Plant and Soil*. 119: 127-132
- Stirling, M., Stirling, G. 1997. "Disease Management: Biological Control," in *Plant Pathogens and Plant Diseases*. Eds. Brown J., Ogle H. 427–439.
- Sturz, A.V., Carter, M.R., Johnston, H.W. 1997. A review of plant disease, pathogen interactions and microbial antagonism under conservation tillage in temperate humid agriculture. *Soil and Tillage Research* 41:169-189
- Sundheim, L. and Tronsmo, A. 1988. Hyperparasites in biological control. In: *Biocontrol of Plant Diseases*. K. G. Mukerji, and K. L. Garg, eds., CRC Press, Boca Raton, FL. 53-69.
- Sylvia, D.M., Hartel, P.G., Fuhrmann, J.J., Zuberer, D.A. 2005. In: Sylvia DM, editor. *Principles and Applications of Soil Microbiology* (2nd Ed.). Upper Saddle River, New Jersey: Pearson Prentice Hall.
- Zhang, Q., Zhang, J., Yang, L., Zhang, L., Jiang, D., Chen, W. 2014. Diversity and biocontrol potential of endophytic fungi in *Brassica napus*. *Biol. Control*. 72: 98–108.